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Note

A method for estimating deuterium oxide density gradients from shifts in the glass-electrode potential

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In a previous paper from this laboratory¹, the utility of deuterium oxide (D_2O) as a density-gradient solute in small isoelectrofocusing columns was demonstrated. A D_2O density gradient created directly in a 1.5-ml column by free interdiffusion of three D_2O solutions for 3 min (ref. 2) was sufficiently strong and sufficiently stable with time to stabilize protein zones of the usual concentrations against convection during isoelectric focusing.

Density gradients made from D_2O have also proved useful for the separation of biological species by centrifugation. According to a recent paper by Trinick and Rowe³, zonal-velocity sedimentation in such gradients seems to be the only known successful way of fractionating thick filaments from vertebrate skeletal muscle.

In work with H_2O-D_2O gradients, it is generally of interest to know the initial density course in the gradient and its stability with time. Previously¹, I obtained this information by *in situ* measurement of the refractive-index gradient as described by Rilbe and Pettersson², whereas Trinick and Rowe³ evaluated the density gradient by pycnometry of the fractions. It is a unique feature of the former technique that the H_2O-D_2O gradient may be repeatedly recorded at desired intervals of time. On the other hand, it is a serious drawback that apparatus for direct photography of refractive-index gradients is nowadays available only to a limited number of biochemical laboratories.

The pycnometer method has proved useful also for very small samples⁴. It is self-evident, however, that accurate evaluation of the density of a H_2O-D_2O fraction containing only 0.06 ml (as in ref. 1) is a difficult task, requiring not only a sensitive balance but also manual skill and patience. Overflow will be an additional complication if the filling of the micro-pycnometer has to be made at a temperature lower than room temperature (*c.g.* at 5° as in the cited works).

The present paper suggests an alternative method for evaluating D_2O density gradients; this method is based on the shift in the asymmetry potential of a glass electrode in D_2O as compared with H_2O .

PRINCIPLE OF THE METHOD

Several workers have reported that, when a glass electrode is used to measure the acidity of a D_2O solution at 25°, the observed pH-meter reading is about 0.4 pH

unit lower than for a H₂O solution of equal acidity⁵. For mixtures of H₂O and D₂O, the deviation in pH-meter reading, $\angle 1p(DH)^*$, is almost proportional to the atom fraction, *n*, of deuterium in the mixture. This is evident from pH-meter readings made by Glasoe and Long⁶, and by Salomaa *et al.*⁷, on solutions containing the same amount of strong acid (*e.g.*, 0.01 *M* hydrochloric acid) but different amounts of D₂O. In principle, therefore, it should be possible to estimate the value of *n* (and hence the density) of an unknown H₂O-D₂O mixture by making the mixture (or a sample of it) 0.01 *M* in hydrochloric acid and reading its "pH". A comparison with the pH-meter reading of D₂O-free 0.01 *M* hydrochloric acid and a calibration curve of $\angle 1p(DH)$ *vs. n* would then give the value of *n*.

EXPERIMENTAL

A series of D_2O density gradients was prepared in the 1.5-ml column described elsewhere^{1.8,9} by free interdiffusion of three aqueous solutions that contained 0.0,50.0 and 97.5% of D_2O by volume as previously¹, but, in addition, were each 0.01 *M* in hydrochloric acid. The solutions were prepared by adding, from a constriction pipette, 0.200 ml of 0.5 *M* hydrochloric acid in H₂O to 9.8 ml of either H₂O or H₂O mixed with D_2O in the appropriate volume ratio. The resulting repeatability in acidity as obtained from pH-meter readings on three preparations of the D_2O -free 0.01 *M* hydrochloric acid was \pm 0.004 pH unit.

The prepared density gradient was left in the column (kept in upright position) for 2 h, then the column contents were fractionated into twenty-five 0.06-ml fractions¹⁰, which were subjected to pH analysis.

The pH measurements were made with a micro electrode unit (type E 5021; Radiometer, Copenhagen, Denmark) coupled to a precision pH meter with a built-in 10-fold scale expander (Radiometer, type PHM26). The glass electrode (G297/G2) in the micro electrode unit was of the capillary type and required about 0.02 ml of sample to give a pH reading. The temperature of the water-jacketed electrodes was kept constant at 25.0°. The pH meter was standardized with conventional buffer solutions of pH 2 and pH 4.

The pH measurements were started by filling the glass capillary several times with the D₂O-free initial solution. (Henceforth, the constant value thus obtained will be denoted by pH₀.) Then the capillary was filled twice with each of the 25 fractions in order of increasing D₂O content, and the pH-meter reading was taken 1 min after the filling. Finally, additional readings were made on the D₂O-free hydrochloric acid. In conformity with earlier observations¹, the first of the additional readings was about 0.02 pH unit higher than pH₀, whereas subsequent ones slowly decreased to pH₀. Since the observed positive shift (δ) in pH-meter reading is probably due to a successive penetration of D₂O into the glass membrane, and since the fractions were measured in the order of increasing D₂O concentration with no rinse in between, a negative correction equal to 0.04 δ multiplied by the fraction number was applied on the second pH-meter reading for each fraction. The corrected reading is denoted by pH_f (f for fraction).

^{*} The value of dp(DH) is defined as the meter reading in H₂O minus the reading in the mixed solvent.

The volume fraction, x_v , of D_2O in the solvent of each fraction was calculated from the difference $pH_0 - pH_f = \Delta p(DH)$ by means of the empirical equation:

$$\Delta p(DH) = 0.325 x_V + 0.060 x_V^2 \tag{1}$$

This equation was derived by the method of least squares from pH-meter readings made earlier¹ on ten solutions of hydrochloric acid prepared as described above and containing 0-90% of D_2O .

RESULTS AND DISCUSSION

The volume percentages of $D_2O(100x_v)$ resulting from two identical densitygradient preparations are plotted in Fig. 1 vs. fraction number and vs. column level. For comparison, Fig. 1 also contains the corresponding concentration course as obtained by measurement of the refractive-index gradient (*i.e.*, the curve marked \bullet ---- \bullet in Fig. 2 of ref. 1).

The density scale of Fig. 1 was calculated from the D₂O-concentration scale as follows. Since mixtures of H₂O and D₂O are ideal, their densities (ϱ) vary linearly with x_{ν} . At 5°, the densities of pure H₂O and pure D₂O are 1.000 and 1.106 g/ml, respectively¹¹; this gives the relation ϱ (g/ml) = 1.000 + 0.106 x_{ν} .

It can be seen from Fig. 1 that the concentration courses estimated by the

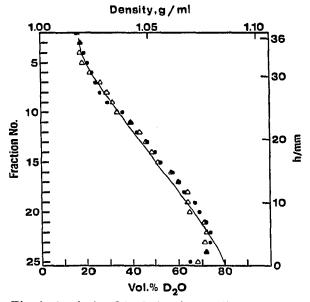


Fig. 1. Analysis of D_2O density gradients prepared at 5° in a 1.5-ml column by free interdiffusion² of three initial solutions containing 0.0, 50.0 and 97.5% of D_2O by volume, respectively. The solid curve was obtained by measurement of the refractive-index gradient². The filled circles and the triangles refer to the D_2O concentrations of 0.06-ml fractions from separate experiments and are derived by comparing pH-meter readings with a calibration curve (for details see text). The prepared density gradients had been left in the column (kept in the vertical position) for 2 h before the analysis. The density scale was calculated from the D_2O -concentration scale by means of the relation $\varrho(g/ml) = 1.000 + 0.106 x_V$ (see text). The vertical distance from the bottom of column is indicated by h.

method described agree well with the one evaluated by refractometry. The deviation is less than ± 4 vol. % of D₂O except for fractions No. 24 and 25. The concentration values obtained for these fractions are not relevant, however, as the fractions were contaminated by the D₂O- and hydrochloric acid-free sucrose solution¹⁰ used for displacement of the column contents at the fractionation.

If the regression straight lines are calculated from the resulting densities of fractions No. 3-22 as given in Fig. 1, density gradients of 0.0218 g/ml·cm (filled circles) and 0.0211 g/ml·cm (triangles) are obtained. The corresponding density gradient pertinent to the solid curve shown in Fig. 1 is 0.0206 g/ml·cm. Thus, both the density-gradient values estimated by "pH" measurements are higher than the value evaluated by refractometry (by 6 and 2.5 %, respectively). This result may be due to an inaccuracy in the solid curve of Fig. 1, but it is also conceivable that eqn. 1 is not pertinent (*cf.* below) or that the corrections applied to the pH-meter readings to give pH_f are not adequate. In any event, the observed uncertainty in the density-gradient strength should normally be insignificant when D₂O density gradients are being used for stabilization of isoelectrofocusing systems or for preparative sedimentation runs.

Likewise, the observed uncertainty in the D_2O concentrations estimated by the present method should normally be unimportant if the goal is to evaluate isoelectric points from runs in D_2O density gradients. The "absolute" p/ value that can be calculated¹ for a protein component focused in such a gradient is certainly influenced by the value of x_V assigned to the focusing level, as the former value is obtained by adding the pertinent value of $\Delta p(DH)$ to the apparent p/ primarily evaluated from the measured pH course. However, according to eqn. 1, an assumed error of ± 0.04 in x_V would induce an error of only ± 0.013 pH unit in $\Delta p(DH)$.

The potential accuracy of the present method of estimating D_2O concentrations is dependent on the validity of the determined values of $\angle lp(DH)$ and of the equation used for calculating x_{ν} from these values. As was mentioned earlier, the $\angle lp(DH)$ value corresponding to 100 vol. % of D_2O is about 0.4 pH unit, which means that the average increase in $\angle lp(DH)$ is only 0.004 pH unit per 1 vol. % of D_2O . Consequently, a precision pH meter is required to obtain sufficient accuracy in pH₀ and pH_f. However, it is also important to control accurately the temperature of the pH cell by means of a thermostat. Around 25°, the pH-meter reading for 0.01 *M* hydrochloric acid decreased about 0.02 pH unit as the temperature of the cell was raised by 1°.

The equation used here for calculating x_V from measured values of $\Delta p(DH)$ may be compared with the data of Glasoe and Long, and of Salomaa *et al.* If the pH-meter readings for 0.01 *M* hydrochloric acid solutions given by Glasoe and Long in Fig. 1 of their paper⁶ are transformed into an equation by the method of least squares, the following relation is obtained:

$$\Delta p(DH) = 0.319x_V + 0.072x_V^2$$
⁽²⁾

In the interval $0 \le x_{\nu} \le 0.8$, the $\Delta p(DH)$ values obtained from eqns. 1 and 2 for a given value of x_{ν} , agree to within ± 0.002 pH unit.

The equation proposed by Salomaa $et al.^7$

$$\Delta p(DH) = 0.3317n + 0.0766n^2$$
(3)

gives values of $\Delta p(DH)$ that are always higher than those obtained from eqn. 1, the difference increasing as *n* increases. It should be noted, however, that, if eqn. 3 is used instead of eqn. 1 to evaluate the percentages (by volume) of D_2O in fractions No. 3-22 in Fig. 1, agreement with the solid curve is considerably better. For example, the arithmetic mean of the deviation from the said curve is -0.4 vol.% if eqn. 3 is used, whereas the corresponding figure for eqn. 1 is +3.1 vol.%.

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REFERENCES

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- 1 S. Fredriksson, J. Chromatogr., 108 (1975) 153.
- 2 H. Rilbe and S. Pettersson, Separ. Sci., 3 (1968) 535.
- 3 J. A. Trinick and A. J. Rowe, *MSE Application Information*, A9, Measuring & Scientific Equipment, Crawley, Great Britain, 1973.
- 4 J. K. Taylor, in I. M. Kolthoff, P. J. Elving and E. B. Sandell (Editors), *Treatise on Analytical Chemistry*, Part I, Vol. 7, Wiley, New York, p. 4584.
- 5 A. K. Covington, M. Paabo, R. A. Robinson, and R. G. Bates, Anal. Chem., 40 (1968) 700; and references therein.
- 6 P. K. Glasoe and F. A. Long, J. Phys. Chem., 64 (1960) 188.
- 7 P. Salomaa, L. L. Schaleger and F. A. Long, J. Amer. Chem. Soc., 86 (1964) 1.
- 8 M. Jonsson, S. Pettersson and H. Rilbe, Anal. Biochem., 51 (1973) 557.

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- 9 H. Rilbe, Ann. N.Y. Acad. Sci., 209 (1973) 80.
- 10 S. Fredriksson, Anal. Biochem., 50 (1972) 575.
- 11 Landolt-Börnstein, Zahlenwerte und Funktionen, Vol. II, Part 1, Springer, Berlin, 6th ed., 1971, pp. 36 and 853.